

<b>JDM MUSCLE BIOPSY INSTRUCTIONS</b> For Use BY OUTSIDE CENTRES UK	<b>DATE FORM COMPLETED</b>
--	----------------------------

<b>Patient R and R Number</b>		<b>Date of Birth (d/m/y)</b>	
<b>Date of MUSCLE biopsy (d/m/y)</b>		<b>Length of symptoms before biopsy</b>	
<b>Date of onset of JDM</b>		<b>COMMENTS</b>	
<b>Please answer the following questions:</b>			
1) Was the patient on steroids before, <b>or</b> at the time that the biopsy was taken?	<input type="checkbox"/> <b>Yes</b> <input type="checkbox"/> <b>No</b>		
a) If yes specify for how many months the patient was on steroids			
2) Was the patient on any DMARDs before, <b>or</b> at the time that the biopsy was taken?	<input type="checkbox"/> <b>Yes</b> <input type="checkbox"/> <b>No</b>		
a) If yes specify for how many months the patient was on DMARDs			
b) If yes specify list of DMARDs			
<b>Muscle biopsy written report from pathologist</b>	If possible paste in or attach here		
<b>Please provide contact details of histopathologist including e mail and telephone</b>	<b>Name</b>	<b>Contact info</b>	
<b>Print Name of Consultant</b>	<b>Date (d/m/y)</b>		

<b>JDM MUSCLE BIOPSY INSTRUCTIONS</b>	<b>DATE FORM COMPLETED -</b>
---------------------------------------	------------------------------

<b>Patient R and R Number</b>	
-------------------------------	--

**INSTRUCTIONS TO SEND BIOPSY SLIDES TO LONDON**

**Protocol for slides preparation**

**A. Routine stains**

Each centre is asked to provide one each of the following stains, which were prepared at the time of diagnostic stain work in their own department:

- H and E
- Trichrome
- ATPase pH 4.6 (or as performed routinely in your lab)
- ATPase pH 9.4 (or as performed routinely in your lab)
- Acid phosphatase
- NADH

Each of these stains should be done in duplicate in the local centre/ hospital, so that one could be kept in the department and the other sent for the study. Consent in the R and R covers this. These sections should be completed and sent at room temperature in one box marked A and labelled with patient study code .

**B sections for immuno histochemistry**

1. Embed section in a cross-sectional orientation in OCT (embedding medium). Cut sections immediately after mounting. Keep block at -20<sup>0</sup>C or below during cutting ; for long term storage keep at -80<sup>0</sup>C
2. Using a cryostat set at -20<sup>0</sup>C or below, cut 15-20 sections 7µm (microns) thick, and transfer to polylysine coated slide. Please try to transfer sections onto slide in the same orientation. Place 2 sections per slide, approximately 2cm apart from each other. Label slides with date, patient study code and a serial number ie 1,2,3 etc....
3. Allow slides to air dry at room temperature overnight, or for a minimum of 2hr.
4. Fix sections in 100% fresh Acetone at room temperature for 10 minutes
5. Allow to air dry, store in carriage boxes (B, C, etc) at room temperature
6. These 15 slides will be transported in boxes (up to 5 slides per box, 2 sections per slide) marked B, C etc .  
**DO NOT RE FREEZE THESE SECTIONS BEFORE POSTING**
7. **TRANSPORT to destination** : all boxes of slides (A, B, C etc) should be transported at **room temperature** and must reach destination **within 1 week** of the day that they were cut.

<b>Please add any other comments:</b>	
---------------------------------------	--

		Please print and sign form
<b>Print Name of Consultant</b>	<b>Date (d/m/y)</b>	